Computers in Biology and Medicine COVID-19 Detection from Mass Spectra of Exhaled Breath --Manuscript Draft--

Manuscript Number: Full Length Article Article Type: Keywords: SARS-CoV-2; COVID-19; Artificial Intelligence; MachineLearning; Data Mining; Breathomics; Mass Spectrometry; MEMS; NEMS Corresponding Author: Giovanni Squillero Polytechnic of Turin Torino, Piemonte ITALY First Author: Nicolò Bellarmino Order of Authors: Nicolò Bellarmino Giorgio Bozzini Riccardo Cantoro Francesco Castelletti Michele Castelluzzo Carla Ciricugno Raffaele Correale Daniela Dalla Gasperina Francesco Dentali Giovanni Poggialini Piergiorgio Salerno Giovanni Squillero Stefano Taborelli Abstract: According to the World Health Organization, the SARS-CoV-2 virus generated a global emergency between 2020 and 2023 resulting in about 7 million deaths out of more than 750 million individuals diagnosed with COVID-19. During these years, polymerase-chain-reaction and antigen testing played a prominent role in disease control. In this study, we propose a fast and non-invasive detection system exploiting a proprietary mass spectrometer to measure ions in exhaled breath. We demonstrated that infected individuals, even if asymptomatic, exhibit characteristics in the air expelled from the lungs that can be detected by a nanotech-based technology and then recognized by soft-computing algorithms. A clinical trial was ran on about 300 patients: the mass spectra in the 10-351 mass-to-charge range were measured, suitably preprocessed, and analyzed by different classification models; eventually, the system shown an accuracy of 95% and a recall of 94% in identifying cases of COVID-19. With performances comparable to traditional methodologies, the proposed system could play a significant role in both routine examination for common diseases and emergency response for new epidemics. Suggested Reviewers: Opposed Reviewers:

Declaration of interests

⊠The authors declare that they have no known	competing financial	interests or personal	relationships
that could have appeared to influence the work	k reported in this pap	oer.	

 \Box The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

COVID-19 Detection from Mass Spectra of Exhaled Breath

Nicolò Bellarmino^a, Giorgio Bozzini^d, Riccardo Cantoro^a, Francesco Castelletti^c, Michele Castelluzzo^b, Carla Ciricugno^b, Raffaele Correale^b, Daniela Dalla Gasperina^c, Francesco Dentali^c, Giovanni Poggialini^c, Piergiorgio Salerno^c, Giovanni Squillero^{a,*}, Stefano Taborelli^c

^aPolitecnico di Torino, Torino, Italy ^bNanoTech Analysis Srl, Torino, Italy ^cUniversità degli Studi dell'Insubria, Varese, Italy ^dHospital ASST Lariana, Como, Italy

Abstract

According to the World Health Organization, the SARS-CoV-2 virus generated a global emergency between 2020 and 2023 resulting in about 7 million deaths out of more than 750 million individuals diagnosed with COVID-19. During these years, polymerase-chain-reaction and antigen testing played a

^{*}Corresponding author.

Email addresses: nicolo.bellarmino@polito.it (Nicolò Bellarmino), giorgio.bozzini@asst-lariana.it (Giorgio Bozzini), riccardo.cantoro@polito.it (Riccardo Cantoro), fcastelletti@studenti.uninsubria.it (Francesco Castelletti), m.castelluzzo@nanotechanalysis.com (Michele Castelluzzo), c.ciricugno@nanotechanalysis.com (Carla Ciricugno), r.correale@nanotechanalysis.com (Raffaele Correale), d.dallagasperina@uninsubria.it (Daniela Dalla Gasperina), Francesco.Dentali@asst-settelaghi.it (Francesco Dentali), giovanni.poggialini@asst-settelaghi.it (Giovanni Poggialini), piergiorgio.salerno@asst-settelaghi.it (Piergiorgio Salerno), giovanni.squillero@polito.it (Giovanni Squillero), stefano.taborelli@asst-settelaghi.it (Stefano Taborelli)

Authors are listed in alphabetical order.

prominent role in disease control. In this study, we propose a fast and non-invasive detection system exploiting a proprietary mass spectrometer to measure ions in exhaled breath. We demonstrated that infected individuals, even if asymptomatic, exhibit characteristics in the air expelled from the lungs that can be detected by a nanotech-based technology and then recognized by soft-computing algorithms. A clinical trial was ran on about 300 patients: the mass spectra in the 10–351 mass-to-charge range were measured, suitably pre-processed, and analyzed by different classification models; eventually, the system shown an accuracy of 95% and a recall of 94% in identifying cases of COVID-19. With performances comparable to traditional methodologies, the proposed system could play a significant role in both routine examination for common diseases and emergency response for new epidemics.

Keywords: SARS-CoV-2, COVID-19, Artificial Intelligence, Machine Learning, Data Mining, Breathomics, Mass Spectrometry, MEMS, NEMS

1. Introduction

The World Health Organization (WHO) reports that the COVID-19 outbreak spreaded all over the world in 2019 caused more than 750 million infections and slightly less than 7 million deaths¹. Unprecedented efforts have been made to reduce the rate of infection, including social restriction and testing [1, 2].

Real-time quantitative polymerase chain reaction (RT-qPCR) has been widely used for identifying infected subjects and illness management. A test

¹https://covid19.who.int/ (retrieved on May 31, 2023)

using this technology can identify SARS-CoV-2 ribonucleic acid on nasopharyngeal or oropharyngeal swabs [3, 4]. However, due to its high sensitivity, reliable results require solid experimental design and an in-depth understanding of normalizing procedures are required [5]; false negative, that is, infected subjects reported as healthy, may occur for several of reasons, such as technical variables in sample collecting, transporting, and handling of viral RNA, genetic diversity, sample types, viral load, and viral exposure time [6]. Moreover, due to the requirement of authorized laboratories with a minimum biosafety level 2 (BSL-2), shipping, processing, and reporting of samples could overload laboratory facilities and delay the test results [7], not to consider the expensive equipment and reagents.

Alternative strategies have been proposed trying to provide tests that are at the same time rapid, not expensive, easy to use, and allowing the identification of infections at earlier stages. Exhaled breath contains respiratory droplets and other small molecules, products of metabolic and catabolic activities, that have been used as indicators of several diseases like lung diseases, breast cancer, diabetes, and other infectious diseases as influenza. The possibility to extend their use to detect COVID-19 provides several advantages over more traditional methods [8, 9]: breath analysis is non-invasive, which means that it does not require a healthcare provider to collect a sample using a swab or other invasive procedure that may result in uncomfortable situations for the patient under testing; breath analysis can be performed rapidly, which means that test results can be obtained quickly allowing early detection and preventing the spread of the virus; finally, breath analysis is a relatively inexpensive and portable testing method, which means that not

only hospitals and clinics, but also airports or other densely crowded locations where real-time testing may be of use.

Furthermore, differently from the simple virus presence, breath analysis could potentially provide information about the stage of infection. For example, certain patterns of volatile organic compounds (VOCs) in the breath may be more common in the early stages of infection, while other patterns may be more common in the later stages. This can be useful for determining the appropriate course of treatment for an infected individual. This holds also for the severity of the infection, differentiating between severe and mild infections, and providing useful information in determining the level of care that an infected individual may require [10].

In this paper, we propose a detection system that uses mass spectrometry and soft computing to quickly analyze exhaled breath from patients and identify the presence of COVID-19. The approach does not require the prior identification of VOCs, but rather relies directly on the analysis of the mass spectrum. We exploit a proprietary nano-sampling device paired with a high-precision mass spectrometer able to perform mass spectra analysis in the range 10 m/z to 351 m/z almost in real time (i. e., usually in few seconds and in less than few minutes in the most unfavorable cases). The raw data are filtered and pre-processed to extract relevant information, then analyzed using standard machine learning (ML) classifiers to detect the COVID-19 presence. The proposed system does not need any reagent and does not produce any hazardous waste; breath samples could be preserved in special containers, allowing the collection to be easily performed by non-specialized staff in different locations. The clinical evaluation performed on about 300

volunteers demonstrates these claims and its reliable performances (namely, 95% accuracy, 94% recall, 96% specificity, and F1-score of 92%).

The rest of the paper is organized as follows: Section 2 introduces the problem; Section 3 sketches the proposed approaches; and Section 4 reports the experimental evaluation. Section 5 concludes the paper.

2. Background and Related Work

The biological markers, or simply biomarkers, are measurable indicators of some biological state or condition. They are medical signs, or objective indications of medical state observed from outside the patient with accurate and reproducible measurements. In biomedicine, the research of disease-specific biomarkers has grown in importance. Finding dysfunctional healthy states in patients, possibly in the early stages of the disease, is the aim of biomarker discovery.

The metabolome of a biological system refers to its set of metabolites with a molecular weight of less than 2000 Da. VOCs are a sub-category of this set [11] that includes small molecules with weight less than 500 Da [12]; such compounds, that include hydrocarbons, alcohols, ketones, aldehydes, and esters, can be directly measured in the gas phase, even through non-invasive and online monitoring [13].

Breath metabolomics, or *breathomics*, provides insight into all the metabolic processes in the body [10]. Human exhaled breath contains respiratory droplets and VOCs [14] produced by metabolic and catabolic human activities [15]. These molecules can be used as indicators for lung diseases [16], breast cancer and diabetes, or other infectious diseases as influenza

[17, 18, 19, 20]. Exhaled VOCs offer a singular source of beneficial biomarkers that are directly correlated with the body's metabolism [21, 22], phenotype and health status [15, 23]. The diseases' chemical patterns, namely its *volatilome*, allow for the evaluation at early stages even spreading diseases, like COVID-19 [24].

Breath analysis has the potential to expand the range of diagnosis platforms for fast and accurate detection of a disease at an early stage, or for metabolic phenotyping, and so contributing to the development of precision medicine and treatment optimization [10, 25]: it is noninvasive, convenient, and fast and several different technologies and approaches are being used to develop breath analysis devices for the detection of COVID-19 and other diseases [26, 27, 28]. Among the many approaches, mass spectrometry can be used to measure the chemical composition of a person's breath, and machine learning algorithms can be trained to recognize patterns associated with specific diseases.

Mass spectrometry (MS) has become an essential tool in the detection and quantification of biomolecules. Its applications in the clinical area, mainly due to high precision and sensitivity, have allowed the discovery of molecular signatures related to SARS-CoV-2 infection and proved useful to select the therapeutic approach and monitoring the patient [29].

MS is an analytical technique used to identify and characterize chemical compounds based on their mass-to-charge ratio (m/z). This technique offers high sensitivity [30] and finds use across various disciplines, including proteomics [31], metabolomics [32], pharmacology [33], and environmental science [34].

At its core, mass spectrometry involves the separation and analysis of ionized molecules based on their mass-to-charge ratio. The first step in mass spectrometry is ionization, where the analyte molecules are converted into gas-phase ions. Once ionized, the ions are separated based on their m/z ratio through a mass analyzer. A common choice among mass analyzers is the quadrupole, which uses a combination of direct current and radiofrequency (RF) fields to selectively transmit ions based on their m/z ratio, in high vacuum conditions. Vacuum pumps are essential for creating and maintaining a high vacuum environment within the mass spectrometer, enabling efficient ion transport that ensures high sensitivity of the instrument. Finally, a detector measures the ion currents and convert them into electrical signals that are processed to produce a mass spectrum, which represents the abundance of ions at different m/z values.

3. Proposed Approach

3.1. Proprietary Mass-Spectra Technology

Standard mass spectrometers are subject to stringent constraints in terms of inlet sample flows and vacuum conditions. Obtaining high-sensitivity requires for the gas to be in a molecular regime inside the MS device. This condition ensures reduced scattering effects, and consequent losses, in the ion beam. The condition of molecular regime imposes requirements on the geometrical dimension of the system and, ultimately, on the pressure in the ionization chamber and the pressure in the MS sensing chamber. Given standard MS inlet fluxes, the pressure requirements to obtain a molecular regime can only be achieved by complex differential vacuum systems, bulky

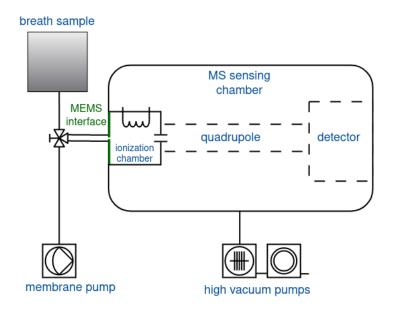


Figure 1: A schematic representation of the experimental apparatus.

connections, and expensive vacuum pumps [34].

To get around this problem, a different approach is made possible by a proprietary nano and micro electro-mechanical systems (NEMS/MEMS) [35, 36, 37]. In this specific case, we use a MEMS that is made up of a micrometer-scale orifice [38]. This device is used as interface between the breath sample, at atmospheric pressure, and the ionization chamber, at vacuum conditions. The selected micro-orifices diameters ensure the flux entering the ionization chamber is approximately at molecular regime [39, 34], even at high gradient pressure. Since also the exiting flux to the quadrupole is at molecular regime, due to the high vacuum conditions in the MS sensing chamber, the same concentrations of the sampled gas mixture in the tedlar bag are kept into the ionization chamber, but at lower pressure. Figure 1 shows a schematic representation of the experimental setup.

This device allows to reduce the inlet gas flow by more than four orders of magnitude, with respect to standard MS gas analyzers, and is able to operate with ultra-simplified vacuum systems [34]. Furthermore, this new technology allows to sample at constant pressure instead of constant flux, removing the need for additional tools for flux control [34].

Such devices allow to optimize sensitivity and to reduce measurement time, thus providing an ideal tool for several applications, including diagnosis through breath analysis. By studying the mass spectrum, it is possible to obtain information about the chemical composition of the breath, including the presence of various VOCs.

3.2. Pre-processing

Once the obtained dataset composed by the acquisitions of the spectra for each patient has been stored, signals are cleaned through a pre-processing procedure that reduces noise and machine-variation of the acquisitions.

For each acquisition, the recorded m/z positions may be shifted with a specific alignment when the machine records the quantity of the ionized molecules due to measurement noise. A peak-alignment procedure is thus necessary. This procedure enables reducing the noise of the machine and compacting information. The peak alignment procedure is based on moving the peak to the nearest integer position, using them as anchors. The curves between two nearby peaks are stretched or compressed to sustain their original shape, preventing information loss. Peaks with mass i are thus moved to the nearest integer position. Stretching and compression between peaks is done by linear interpolation to fit the corresponding segments in the reference. A graphical plot after the peak alignment can be seen in Figure 3. By

summing all the intensities for each m/z in each acquisition, we can obtain the Total Ion Current (TIC) curve plot in Figure 2. After a certain time, the TIC curve stabilizes, reaching a plateau.

For each patient, we can obtain a single robust mass-spectra measurement by averaging the acquisitions in the plateau zone. To achieve this, first, it is necessary to identify the plateau in the TIC curve by means of gradient computation of the signal. A plateau-searching procedure was implemented, based on looking for acquisitions that do not vary so much from each other. The plateau searching procedure was implemented as follows:

- For each acquisition, we first applied a 1D-uniform filter to smooth the signals. This is based on taking the arithmetic average of each mass intensity value with its neighbors, with a window size of 1.
- We computed the gradient of the signals.
- A plateau is a zone that is almost flat, ideally where the gradient is zero, or in which the gradient does not vary so much from zero: we computed a tolerance guard-band ε, to identify as "flat" a zone in which the gradient is near to zero (i.e., its absolute value is below ε). This ε was computed on the basis of the q th quantile, in which q is a number in the range [0, 1] that indicates how tolerant the requirement of the constant slope of the plateau is.
- Once the plateau of maximum length is found (that vary from 3 to 5 acquisition, for each patient), we computed standard deviations of acquisition in the plateau by deploying a rolling window of size 4. We then found the acquisitions for which the rolling window of standard

deviation is minimum, and we computed the mean among these acquisitions, obtaining a single spectrum for each patient.

The model is trained on all the acquisitions extracted in the plateau zone by the procedure described above. This permits increasing both the number of training samples by a factor of 3-4 and the variability in the data, thus, leading to more accurate models. During the testing phase, instead, we considered a single mass spectrum for each patient, extracted by averaging the multiple acquisitions of each patient.

To eliminate both noise in the measurements and the possible parameters variations in the setting of the machine on different days, a signal smoothing procedure was implemented. We first normalized each spectra by dividing for the TIC, in order to obtain relative information about the breath composition (i. e., each intensity was divided by the sum of all the intensities, thus scaling the feature in the range (0,1)). We then applied a high-pass filter, considering as noise (and thus, treating as zero) each intensity below 0.0001. Then, we applied a Savitzky-Golay Smoothing and Differentiation Filter [40, 41] to remove noise and align the signals to the baseline. This type of filter is used as a pre-processing step in spectra analysis to reduce both high-frequency noises, due to its smoothing properties, and low-frequency signals using differentiation. We then applied again a high-pass filter, considering as zero each intensity below 0.001, to remove possible artifacts introduced by the filtering procedure.

Once implemented the filtering and pre-processing procedure for a single mass range, we applied it to all the mass ranges and combined the obtained spectra. Since we consider different acquisitions for each mass range, merging them together means considering all the combinations of the different acquisitions for each range. In other words, the expansion lead to a dataset in which each row is a particular combination of the different acquisition of each mass range, for each patient. This procedure is something similar to creating artificial rows-patients, in which each of them varies for one of the four pieces of the spectra. As an additional step, we further normalized the whole spectra by the total sum of the intensity, obtaining only relative information in the range (0, 1).

3.3. Machine-Learning Models

Several machine-learning (ML) techniques have been used by scholars in the context of COVID-19 [42, 15]. We propose to exploit state-of-the-art ML models: K-nearest neighbors (KNN) random forest (RF), logistic regression (LR), gradient boosting (XB), support vector machine (SVM) with *RBF* kernel, and an ensemble made by all the model together, in a hard-voting fashion (Ens). To reduce the feature that ML models should analyze, simplify the model, and avoid overfitting, feature reduction techniques are used.

Features are filtered by removing all the feature that present a variance equal to zero (thus, we removed all the m/z for which no intensities were measured in the filtered dataset). Then they are normalized by means of standard normalization approaches (Standard Scaler). The Standard Scaler scales each feature by subtracting the mean and diving by the variance. However, the outliers have an influence when computing the empirical mean and standard deviation. Therefore, it may not ensure a balanced scaling of features in the presence of outliers. To solve this issue, we propose to use a Robust Scaler: the centering and scaling statistics computed by this type

of normalization approach are based on percentiles and are therefore not influenced by a few very large marginal outliers. This removes the median and scales the data according to the interquartile range (the IQR), which is the range between the 1st quartile (25th percentile) and the 3rd quartile (75th percentile).

Then, we used principal component analysis (PCA) [43, 44] to extract the first 20 principal components of our dataset. But before applying the PCA, a further step of feature selection approach can be performed, to select only the most informative features, the ones the most linked to the target task (in a supervised-feature selection fashion).

For this purpose, we used a Relief based algorithm [45], called SURF*. Relief algorithm use a filter-method approach to feature selection that is notably sensitive to feature interactions. It is based on having a feature score. The scoring involves comparing feature values between pairs of neighboring instances to determine their importance. If a feature value difference is detected in a neighboring instance pair with the same class, the feature score is decreased, which is known as a hit. On the other hand, if a feature value difference is observed in a neighboring instance pair with different class values, the feature score is increased, which is referred to as a miss.

The SURF algorithm is a variant of the Relief algorithm that considers nearest neighbors, including both hits and misses, based on a distance threshold from the target instance. The distance threshold is defined by the average distance between all pairs of instances in the training data. According to research [46], SURF has been found to outperform ReliefF in detecting 2-way interactions. SURF* (or SURF Star) [47] is an extension of the SURF

algorithm that not only considers near neighbors but also far instances in the scoring updates. However, inverted scoring updates are employed for far instance pairs.

3.4. Model Evaluation

Patients have been split into training and test data: the training data are used to create the ML models, while the testing data are used only for evaluation purposes. We evaluated the experiments on 10 different training-test splits, averaging the results to obtain an unbiased estimation of the generalization performances of our models. We used a 10-fold Stratified Cross Validation. Results are presented in terms of famous classification performances: balanced accuracy (A), precision (P), recall (R), and F1-Score (F1).

These metrics are computed on the basis of the number of samples correctly and incorrectly predicted by our model and are based on the concept of true positives, true negatives, false positives, and false negatives. A true positive (TP) is an outcome where the model correctly predicts the positive class. Similarly, a true negative (TN) is an outcome where the model correctly predicts the negative class. A false positive (FP) is an outcome where the model incorrectly predicts the positive class. And a false negative FN is an outcome where the model incorrectly predicts the negative class. The balanced accuracy avoids inflated performance estimates on imbalanced datasets. It is the macro-average of recall scores. Intuitively, the precision is the ability of the classifier not to label as positive a sample that is negative, while recall is the ability of the classifier to find all the positive samples. The F1 score can be interpreted as a harmonic mean of the precision and recall,

where an F1 score reaches its best value at 1 and worst score at 0. The relative contribution of precision and recall to the F1 score are equal. In formula, A, P, R, and F1 are computed as below:

$$A = \frac{1}{2} \left(\frac{TP}{TP + FN} + \frac{TN}{TN + FP} \right) \tag{1}$$

$$P = \frac{TP}{TP + FP} \tag{2}$$

$$R = \frac{TP}{TP + FN} \tag{3}$$

$$F1 = 2\frac{P \cdot R}{P + R} \tag{4}$$

For experiments on the whole mass spectra with the four ranges together, we also computed two additional performance metrics, that are Specificity and the area under the receiver operating characteristic curve (ROC-AUC). While the recall is a measure that evaluates a test's ability to correctly identify unhealthy individuals without a particular condition, specificity carry the same concept but for the healthy patients. It is calculated using the formula $specificity = \frac{TN}{TN+FP}$ Essentially, specificity represents the probability of obtaining a negative test result when a patient is actually healthy. When a test exhibits high specificity, a positive result becomes valuable in confirming the presence of the disease, as the test rarely produces positive outcomes in healthy individuals. If a test has 100% specificity, it will correctly identify all patients without the disease by yielding negative results. Thus, a positive test outcome would definitively indicate the presence of the disease. Another way to interpret specificity is as the recall of the negative class.

A receiver operating characteristic (ROC) curve is a visual representation that showcases the performance of a binary classifier system as the threshold for classification is adjusted. It plots the true positive rate (TPR) against the false positive rate (FPR) at different threshold values. TPR is also referred to as sensitivity, while FPR is the complement of specificity.

The ROC-AUC quantifies the overall performance of the classifier by calculating the area beneath the ROC curve. This single numerical value summarizes the information conveyed by the entire curve.

4. Experimental Evaluation

Breath samples were collected from patients and medical personnel at Varese Hospital (Ospedale di Circolo — Fondazione Macchi, ASST Sette Laghi). The acquisition lasted complexively one year, from March 2021 to March 2022, for a total of 302 tested subjects, some tested more than once to calibrate the system. The mass spectra have been collected with a Varian 1200L mass analyzer, combined with the MEMS interface, as shown in Figure 1. As ground truth to confirm SARS-CoV-2 infection, a RT-qPCR nasopharyngeal swab testing has been performed on all subjects.

4.1. Test protocol

The subject's breath is collected into a sampling tedlar bag with a defined volume of 3 litres, by having the subject blow through a straw directly into the bag. Then, the bag is connected to the inlet valve of the MS apparatus.

As shown in Fig. 1, the inlet valve can have two possible settings: the first setting allows for the sample mixture, at atmospheric pressure, to flow from the bag to the ionization chamber, directly through the MEMS interface; the

second setting connects the MEMS interface to a membrane pump, in order to clean the inlet line, bringing it to vacuum conditions ($\simeq 1 \times 10^{-3}$ mbar). Figure 2 shows the TIC behaviour when the breath sample flows into the MS system: the initial increase is due to the abrupt pressure change at the valve opening and, after a few tens of second, TIC curve shows a plateau region[38] when the flux reaches stability.

Mass spectra are recorded via the Varian 1200L mass analyzer software, which allows to set some acquisition parameters like mass range, acquisition time and electron multiplier (EM) voltage. The latter parameter ultimately sets the detector amplification factor.

We recorded mass spectra in the following ranges:

- 10 m/z to 51 m/z, with an acquisition time of 10 s and EM voltage of 1000 V;
- $49 \,\mathrm{m/z}$ to $151 \,\mathrm{m/z}$, with an acquisition time of $14 \,\mathrm{s}$ and EM voltage of $1800 \,\mathrm{V}$;
- $149\,\mathrm{m/z}$ to $251\,\mathrm{m/z}$, with an acquisition time of $14\,\mathrm{s}$ and EM voltage of $1800\,\mathrm{V}$;
- 249 m/z to 351 m/z, with an acquisition time of 14 s and EM voltage of 1800 V;

To avoid signal saturation, the amplification in the first mass range was reduced due to the presence of the most abundant breath components, namely CO_2 (44 m/z), N_2 (28 m/z) and O_2 (32 m/z).

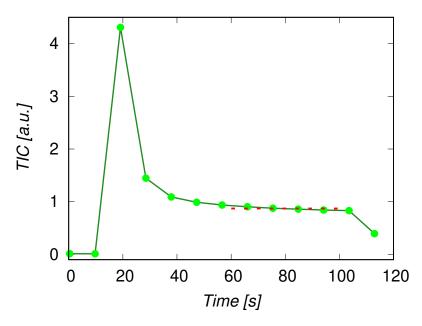


Figure 2: Total Ion Current (TIC) of a recording from one sample. The recording is made up of about 10 acquisitions (green dots), each corresponding to a mass spectrum. The spectra used for the analysis are selected on the plateau of TIC (red dotted region).

4.2. Data Analysis

The raw dataset presents breath samples for a total of 1208 acquisitions in 302 patients, divided into 91 positive records and 211 negative records. After removing outliers (with a z-score greater than 8, for at least one feature) and patient for which it was not possible to identify a plateau, we reached 287 patients for mass range 2 and 203 if we consider all the ranges. These problems were caused by the highly prototypical nature of the equipment; it is worth noticing that, in a real application, it would have been possible to repeat the measurement. To address the issue of high class imbalance, we utilized a simple oversampling technique on the minority class (i. e., the COVID-19 positive class) in each iteration of the training-test procedure.

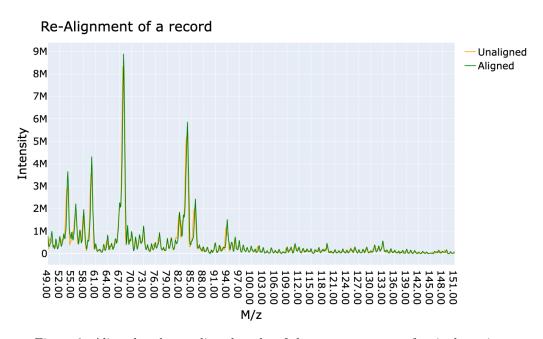


Figure 3: Aligned and non-aligned peaks of the mass spectrum of a single patient.

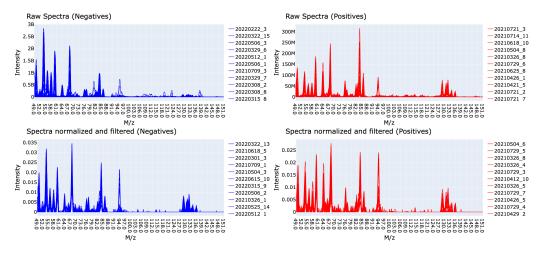


Figure 4: The comparison of spectra before (top) and after the filtering and normalizing procedure (bottom) shows the removal of low-frequency noise. Negative patients are on the left (blue), positive on the right (red).

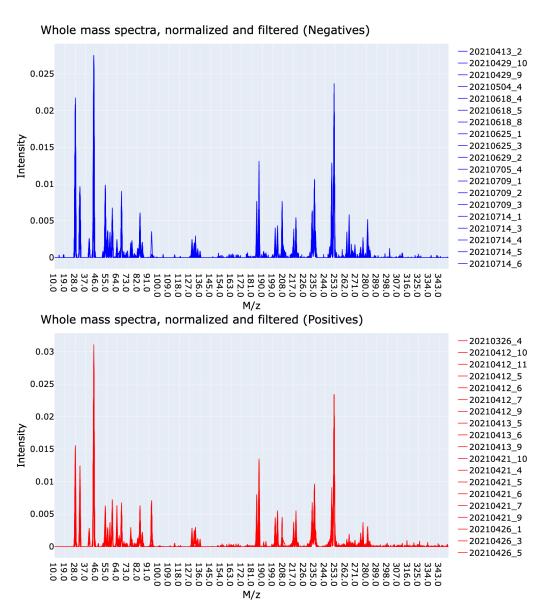


Figure 5: Examples of whole spectra $(10\,\mathrm{m/z}$ to $351\,\mathrm{m/z})$ for negative subjects (top, blue) and positive ones (bottom, red)

2D Representation

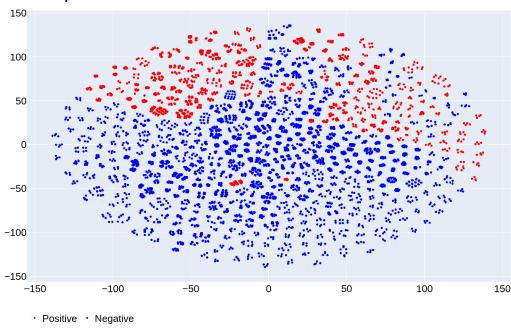


Figure 6: 2D t-SNE representation of the whole spectra for all the 47,084 samples generated.

The results of the approaches described in 3.2 can be found in Table 2. The expansion procedure increased the dimensionality of the dataset to 47 084 samples but created from only 203 patients. A graphical 2D representation of the points obtained with t-SNE dimensionality reduction [48] is presented in figure Figure 6. There, a precise boundary between positive and negative samples can be seen.

The filtering of features reduced the number of features from 1021 to 611 in the mass range 2 and from 3411 to 1734 in the whole spectrum. The results of training on different mass ranges and testing them on the test sets without feature pre-processing seems to indicate mass range 2 as the more useful for classification with a F1 score of 0.70 and accuracy of 0.83 (Tables 1

and 2).

As shown in Table 2, the idea of inserting multiple acquisition for each patient in the training set, while averaging them on the test set, lead to a lower prediction error and higher classification performance with respect to a single, robust spectrum. With the Ensemble model, we can reach the 91.4% of balanced accuracy and the 86.9% of F1-Score if we include all the stable acquisitions in the training set, while we reached the 89.8% of accuracy and 84.8 % of F1-Score with the same ensemble model, but with a single acquisition per patient. This hold also for the other model taken into account. Filtering and normalizing the spectra is also beneficial, since it permits to increase the classification performance for all the models: if we compare Table 1 and Table 2, it is evident that, for range 2, we increased the accuracy from 84% to 93.4%, and the F1-Score from 70% to about 89%.

The SVC and Ensemble algorithms have the highest recall values for all the data processing setup, indicating that they have the highest ability to correctly identify true positives patients. In general, these two algorithms have the highest prediction performances, that, in principle, could be additionally improved with a fine search in the hyperparameters.

Including a Feature Selection steps was not beneficial in terms of error metrics: with the SURF* algorithm, the error metrics decrease. It may be due to complex interaction between features, that this algorithm is not able to catch. The robust scaler permits to decrease the standard deviation in the performance metrics, and thus the error fluctuations, due to the ability to deal with outliers values. It is extremely beneficial in the case of SVM, and also with the Ensemble method.

Using all the spectra together, and the subsequent artificial samples creation technique permitted used to reach the best performances in terms of error metric Table 3: with the ensemble method, we achieve 95% accuracy, 94% recall, 96% specificity, and F1-score of 92%.

Table 1: Average results with different mass ranges on the test set on 10 runs, no features pre-processing

Model	Mass Range	Accuracy	Precision	Recall	F1-Score
RF	1	0.78	0.65	0.56	0.60
RF	2	0.82	0.69	0.69	0.68
RF	3	0.82	0.72	0.57	0.63
RF	4	0.79	0.65	0.53	0.57
Ens	1	0.82	0.70	0.67	0.68
Ens	2	0.84	0.75	0.69	0.71
Ens	3	0.80	0.68	0.57	0.61
Ens	4	0.77	0.62	0.60	0.59

5. Conclusions

We used a prototype of a special MS portable machine able based on nanotechnology to analyze human breath in about 2 minutes, extracting the mass spectrum of a patient in the interval 10-351, divided into four subranges. Experiments showed that the mass spectra can be related to the presence of COVID-19 by means of ML classification models. We proposed a filtering procedure based on Savitzky-Golay filter to reduce possible noise in the acquisitions. Results showed that with simple approaches we are able to get about 93% accuracy and 94% recall with mass range of 49 m/z to 151 m/z, which seems to be the most appropriate in predicting COVID-19

Table 2: Mean Results of Classification Algorithms on the test sets with mass range 2

Alg.	Filtering	Feat. Sel.	Acquisition	Accuracy	Precision	Recall	F1 Score
xGB	No	PCA	Single	0.83 ± 0.04	0.74 ± 0.10	0.77 ± 0.08	0.75 ± 0.06
xGB	Yes	PCA	Single	0.88 ± 0.07	0.79 ± 0.08	0.86 ± 0.13	0.82 ± 0.09
xGB	Yes	PCA	Multiple	0.93 ± 0.05	0.85 ± 0.14	0.94 ± 0.07	$\textbf{0.88}\pm\textbf{0.09}$
xGB	Yes	SURF*, PCA	Multiple	0.86 ± 0.07	0.78 ± 0.14	0.83 ± 0.10	0.80 ± 0.10
xGB	Yes (RS)	PCA	Multiple	0.90 ± 0.04	0.82 ± 0.10	0.89 ± 0.08	0.84 ± 0.05
KNN	No	PCA	Single	0.89 ± 0.05	0.73 ± 0.09	0.92 ± 0.08	0.81 ± 0.07
KNN	Yes	PCA	Single	0.87 ± 0.05	0.73 ± 0.07	0.89 ± 0.08	0.80 ± 0.07
KNN	Yes	PCA	Multiple	0.91 ± 0.07	0.80 ± 0.14	0.93 ± 0.10	0.85 ± 0.11
KNN	Yes	SURF*, PCA	Multiple	0.85 ± 0.07	0.70 ± 0.13	0.87 ± 0.10	0.77 ± 0.11
KNN	Yes (RS)	PCA	Multiple	0.91 ± 0.05	0.78 ± 0.12	0.94 ± 0.08	0.84 ± 0.08
LR	No	PCA	Single	0.86 ± 0.05	0.71 ± 0.12	0.88 ± 0.07	0.78 ± 0.08
LR	Yes	PCA	Single	0.85 ± 0.06	0.71 ± 0.08	0.85 ± 0.10	0.77 ± 0.07
LR	Yes	PCA	Multiple	0.88 ± 0.06	0.73 ± 0.14	0.92 ± 0.10	0.80 ± 0.10
LR	Yes	SURF*, PCA	Multiple	0.88 ± 0.07	0.74 ± 0.13	0.89 ± 0.10	0.80 ± 0.10
LR	Yes (RS)	PCA	Multiple	0.89 ± 0.05	0.76 ± 0.11	0.91 ± 0.08	0.82 ± 0.08
RF	No	PCA	Single	0.87 ± 0.05	0.79 ± 0.10	0.84 ± 0.09	0.81 ± 0.07
RF	Yes	PCA	Single	0.89 ± 0.04	0.82 ± 0.11	0.87 ± 0.08	0.84 ± 0.05
RF	Yes	PCA	Multiple	0.91 ± 0.07	0.80 ± 0.14	0.92 ± 0.09	0.85 ± 0.10
RF	Yes	SURF*, PCA	Multiple	0.85 ± 0.09	0.75 ± 0.16	0.82 ± 0.13	0.78 ± 0.12
RF	Yes (RS)	PCA	Multiple	0.90 ± 0.03	0.81 ± 0.07	0.90 ± 0.07	0.84 ± 0.04
SVC	No	PCA	Single	0.65 ± 0.07	0.82 ± 0.20	0.32 ± 0.13	0.45 ± 0.16
SVC	Yes	PCA	Single	0.76 ± 0.10	0.82 ± 0.15	0.57 ± 0.17	0.66 ± 0.15
SVC	Yes	PCA	Multiple	0.89 ± 0.05	0.76 ± 0.13	0.91 ± 0.13	0.81 ± 0.07
SVC	Yes	SURF*, PCA	Multiple	0.86 ± 0.08	0.77 ± 0.17	0.83 ± 0.09	0.79 ± 0.12
SVC	Yes (RS)	PCA	Multiple	0.93 ± 0.04	0.85 ± 0.09	$\textbf{0.94}\pm\textbf{0.07}$	0.89 ± 0.06
Ens.	No	PCA	Single	0.87 ± 0.03	0.77 ± 0.10	0.84 ± 0.06	0.80 ± 0.06
Ens.	Yes	PCA	Single	0.90 ± 0.07	0.82 ± 0.11	0.89 ± 0.10	0.85 ± 0.09
Ens.	Yes	PCA	Multiple	0.93 ± 0.07	0.83 ± 0.15	0.94 ± 0.09	0.87 ± 0.11
Ens.	Yes	SURF*, PCA	Multiple	0.89 ± 0.07	0.79 ± 0.16	0.88 ± 0.08	0.82 ± 0.12
Ens.	Yes (RS)	PCA	Multiple	$\textbf{0.92}\pm\textbf{0.04}$	0.81 ± 0.09	$\textbf{0.94}\pm\textbf{0.07}$	0.87 ± 0.06

disease. We found out that the use of Robust scaling techniques, based on the median and IQR scaling, in conjunction with PCA feature extraction were beneficial in predicting COVID-19 diseases. We were able to merge all the spectra ranges, reaching classification performances of about 95% of accuracy and 98% of ROC-AUC score. In general, the special portable MS machine is potentially deployable in COVID-19 hubs to detect in a fast, easy, comfortable way the presence or not of COVID-19. The use of ML to relate mass spectra to special diseases enables early detection, rapid test results, and less risk of infection for healthcare providers.

Table 3: Mean Results (10 Splits) for the whole mass range 10-351.

Alg.	Accuracy	Precision	Recall	F1 Score	Specificity	ROC-AUC
KNN	0.93 ± 0.06	0.87 ± 0.09	0.92 ± 0.09	0.89 ± 0.08	0.94 ± 0.04	0.95 ± 0.04
RF	0.91 ± 0.06	0.88 ± 0.10	0.87 ± 0.12	0.87 ± 0.07	0.95 ± 0.04	0.98 ± 0.03
LR	0.94 ± 0.04	0.84 ± 0.12	0.96 ± 0.07	0.89 ± 0.07	0.93 ± 0.05	0.97 ± 0.04
xGB	0.94 ± 0.03	0.88 ± 0.08	0.93 ± 0.07	0.90 ± 0.03	0.95 ± 0.03	0.98 ± 0.03
SVC	0.93 ± 0.06	0.89 ± 0.09	0.90 ± 0.12	0.88 ± 0.06	0.95 ± 0.04	0.98 ± 0.02
Ens.	0.95 ± 0.04	0.90 ± 0.08	0.94 ± 0.07	0.92 ± 0.05	0.96 ± 0.03	0.98 ± 0.03

Acknowledgement

Authors wish to thank Mattee Serra for his contribution.

References

- D. K. Chu, E. A. Akl, S. Duda, K. Solo, S. Yaacoub, H. J. Schünemann, A. El-harakeh, A. Bognanni, T. Lotfi, M. Loeb, A. Hajizadeh, A. Bak, A. Izcovich, C. A. Cuello-Garcia, C. Chen, D. J. Harris, E. Borowiack, F. Chamseddine, F. Schünemann, G. P. Morgano, G. E. U. Muti Schünemann, G. Chen, H. Zhao, I. Neumann, J. Chan, J. Khabsa, L. Hneiny, L. Harrison, M. Smith, N. Rizk, P. Giorgi Rossi, P. Abi-Hanna, R. El-khoury, R. Stalteri, T. Baldeh, T. Piggott, Y. Zhang, Z. Saad, A. Khamis, M. Reinap, Physical distancing, face masks, and eye protection to prevent person-to-person transmission of SARS-CoV-2 and COVID-19: a systematic review and meta-analysis, Lancet (London, England) 395 (10242) (2020) 1973–1987. doi:10.1016/S0140-6736(20)31142-9.
- [2] O. Vandenberg, D. Martiny, O. Rochas, A. van Belkum, Z. Kozlakidis, Considerations for diagnostic COVID-19 tests, Nature Reviews Microbiology 19 (3) (2021) 171–183. doi:10.1038/s41579-020-00461-z.

- [3] W. Feng, A. M. Newbigging, C. Le, B. Pang, H. Peng, Y. Cao, J. Wu, G. Abbas, J. Song, D.-B. Wang, M. Cui, J. Tao, D. L. Tyrrell, X.-E. Zhang, H. Zhang, X. C. Le, Molecular diagnosis of COVID-19: Challenges and research needs, Analytical Chemistry 92 (15) (2020) 10196–10209. doi:10.1021/acs.analchem.0c02060.
- [4] S. Eissa, M. Zourob, Development of a low-cost cotton-tipped electrochemical immunosensor for the detection of SARS-CoV-2, Analytical Chemistry 93 (3) (2021) 1826–1833. doi:10.1021/acs.analchem.0c04719.
- [5] M. L. Wong, J. F. Medrano, Real-time PCR for mRNA quantitation, BioTechniques 39 (1) (2005) 75–85, pMID: 16060372. doi:10.2144/ 05391RV01.
- [6] F. Bahreini, R. Najafi, R. Amini, S. Khazaei, S. Bashirian, Reducing false negative PCR test for COVID-19, International Journal of Maternal and Child Health and AIDS (IJMA) 9 (3) (2020) 408–410. doi:10.21106/ijma.421.
- [7] K. Wang, X. Zhu, J. Xu, Laboratory biosafety considerations of SARS-CoV-2 at biosafety level 2, Health Security 18 (3) (2020) 232–236. doi: 10.1089/hs.2020.0021.
- [8] K. Lamote, E. Janssens, E. Schillebeeckx, T. S. Lapperre, B. Y. De Winter, J. P. van Meerbeeck, The scent of COVID-19: viral (semi-)volatiles as fast diagnostic biomarkers?, Journal of Breath Research 14 (4) (2020) 042001. doi:10.1088/1752-7163/aba105.

- [9] J.-W. Song, S. M. Lam, X. Fan, W.-J. Cao, S.-Y. Wang, H. Tian, G. H. Chua, C. Zhang, F.-P. Meng, Z. Xu, J.-L. Fu, L. Huang, P. Xia, T. Yang, S. Zhang, B. Li, T.-J. Jiang, R. Wang, Z. Wang, M. Shi, J.-Y. Zhang, F.-S. Wang, G. Shui, Omics-driven systems interrogation of metabolic dysregulation in COVID-19 pathogenesis, Cell Metabolism 32 (2) (2020) 188–202.e5. doi:10.1016/j.cmet.2020.06.016.
- [10] N. J. Rattray, Z. Hamrang, D. K. Trivedi, R. Goodacre, S. J. Fowler, Taking your breath away: metabolomics breathes life in to personalized medicine, Trends in Biotechnology 32 (10) (2014) 538–548. doi:https://doi.org/10.1016/j.tibtech.2014.08.003.
- [11] M. Mansurova, B. E. Ebert, L. M. Blank, A. J. Ibáñez, A breath of information: the volatilome, Current Genetics 64 (4) (2018) 959–964. doi:10.1007/s00294-017-0800-x.
- [12] M. Rodríguez-Aguilar, L. Díaz de León-Martínez, B. N. Zamora-Mendoza, A. Comas-García, S. E. Guerra Palomares, C. A. García-Sepúlveda, L. E. Alcántara-Quintana, F. Díaz-Barriga, R. Flores-Ramírez, Comparative analysis of chemical breath-prints through olfactory technology for the discrimination between SARS-CoV-2 infected patients and controls, Clinica Chimica Acta 519 (2021) 126–132. doi:10.1016/j.cca.2021.04.015.
- [13] G. Giovannini, H. Haick, D. Garoli, Detecting COVID-19 from breath:
 A game changer for a big challenge, ACS sensors 6 (4) (2021) 1408–1417.
 doi:10.1021/acssensors.1c00312.

- [14] Use of exhaled breath condensate (ebc) in the diagnosis of SARS-COV-2 (COVID-19) 76 (2021) 86–88. doi:10.1136/thoraxjnl-2020-215705.
- [15] A. Smolinska, A. C. Hauschild, R. R. Fijten, J. W. Dallinga, J. Baumbach, F. J. van Schooten, Current breathomics—a review on data preprocessing techniques and machine learning in metabolomics breath analysis, J Breath Res 8 (2) (2014) 027105. doi:10.1088/1752-7155/8/2/027105.
- [16] R. Gasparri, M. Santonico, C. Valentini, G. Sedda, A. Borri, F. Petrella, P. Maisonneuve, G. Pennazza, A. D'Amico, C. Di Natale, R. Paolesse, L. Spaggiari, Volatile signature for the early diagnosis of lung cancer, Journal of Breath Research 10 (1) (2016) 016007. doi:10.1088/1752-7155/10/1/016007.
- [17] E. Janssens, J. P. van Meerbeeck, K. Lamote, Volatile organic compounds in human matrices as lung cancer biomarkers: a systematic review, Critical Reviews in Oncology/Hematology 153 (2020) 103037—doi:10.1016/j.critrevonc.2020.103037.
- [18] I. Oakley-Girvan, S. W. Davis, Breath based volatile organic compounds in the detection of breast, lung, and colorectal cancers: A systematic review, Cancer Biomarkers 21 (1) (2017) 29–39. doi:10.3233/ CBM-170177.
- [19] B. Behera, R. Joshi, G. K. Anil Vishnu, S. Bhalerao, H. J. Pandya, Electronic nose: a non-invasive technology for breath analysis of diabetes and

- lung cancer patients, Journal of Breath Research 13 (2) (2019) 024001—. doi:10.1088/1752-7163/aafc77.
- [20] A. Abd El Qader, D. Lieberman, Y. Shemer Avni, N. Svobodin, T. Lazarovitch, O. Sagi, Y. Zeiri, Volatile organic compounds generated by cultures of bacteria and viruses associated with respiratory infections: Vocs generated by cultures of bacteria and viruses, Biomedical Chromatography 29 (12) (2015) 1783–1790. doi:10.1002/bmc.3494.
- [21] A. Mazzatenta, C. Di Giulio, M. Pokorski, Pathologies currently identified by exhaled biomarkers, Respiratory Physiology & Neurobiology 187 (1) (2013) 128–134, immunopathology of the Respiratory System. doi:https://doi.org/10.1016/j.resp.2013.02.016.
- [22] B. de Lacy Costello, A. Amann, H. Al-Kateb, C. Flynn, W. Filipiak, T. Khalid, D. Osborne, N. M. Ratcliffe, A review of the volatiles from the healthy human body, Journal of Breath Research 8 (1) (2014) 014001. doi:10.1088/1752-7155/8/1/014001.
- [23] K. Hollywood, D. R. Brison, R. Goodacre, Metabolomics: Current technologies and future trends, PROTEOMICS 6 (17) (2006) 4716–4723. doi:https://doi.org/10.1002/pmic.200600106.
- [24] C. E. Davis, M. Schivo, N. J. Kenyon, A breath of fresh air the potential for COVID-19 breath diagnostics, EBioMedicine 63 (2021) 103183-.
 doi:10.1016/j.ebiom.2020.103183.
- [25] W. Miekisch, J. K. Schubert, G. F. E. Noeldge-Schomburg, Diagnostic potential of breath analysis—focus on volatile organic compounds, Clinica

- Chimica Acta; International Journal of Clinical Chemistry 347 (1–2) (2004) 25–39. doi:10.1016/j.cccn.2004.04.023.
- [26] T. Bruderer, T. Gaisl, M. T. Gaugg, N. Nowak, B. Streckenbach, S. Müller, A. Moeller, M. Kohler, R. Zenobi, On-line analysis of exhaled breath, Chemical Reviews 119 (19) (2019) 10803–10828. doi: 10.1021/acs.chemrev.9b000005.
- [27] A. Z. Berna, E. H. Akaho, R. M. Harris, M. Congdon, E. Korn, S. Neher, M. M'Farrej, J. Burns, A. R. Odom John, Reproducible breath metabolite changes in children with SARS-CoV-2 infection, ACS Infect Dis 7 (9) (2021) 2596–2603. doi:10.1101/2020.12.04.20230755.
- [28] P. Zhang, T. Ren, H. Chen, Q. Li, M. He, Y. Feng, L. Wang, T. Huang, J. Yuan, G. Deng, H. Lu, A feasibility study of COVID-19 detection using breath analysis by high-pressure photon ionization time-of-flight mass spectrometry, Journal of Breath Research 16 (4) (2022) 046009. doi:10.1088/1752-7163/ac8ea1.
- [29] N. M. Lima, B. L. M. Fernandes, G. F. Alves, J. C. Q. de Souza, M. M. Siqueira, M. Patrícia do Nascimento, O. B. O. Moreira, A. Sussulini, M. A. L. de Oliveira, Mass spectrometry applied to diagnosis, prognosis, and therapeutic targets identification for the novel coronavirus SARS-CoV-2: A review, Analytica Chimica Acta 1195 (2022) 339385—. doi: 10.1016/j.aca.2021.339385.
- [30] K. A. Pratt, K. A. Prather, Mass spectrometry of atmospheric aerosols—recent developments and applications. part ii: On-line mass

- spectrometry techniques, Mass Spectrometry Reviews 31 (1) (2012) 17–48. doi:https://doi.org/10.1002/mas.20330.
- [31] R. Aebersold, M. Mann, Mass spectrometry-based proteomics, Nature 422 (6928) (2003) 198 207. doi:10.1038/nature01511.
- [32] K. Dettmer, P. A. Aronov, B. D. Hammock, Mass spectrometry-based metabolomics, Mass Spectrometry Reviews 26 (1) (2007) 51–78. doi: https://doi.org/10.1002/mas.20108.
- [33] D. A. Dias, S. Urban, U. Roessner, A historical overview of natural products in drug discovery, Metabolites 2 (2) (2012) 303–336. doi: 10.3390/metabo2020303.
- [34] L. Franceschelli, C. Ciricugno, M. Di Lorenzo, A. Romani, A. Berardinelli, M. Tartagni, R. Correale, Real-time gas mass spectroscopy by multivariate analysis, Scientific Reports 13 (1) (2023). doi:10.1038/s41598-023-33188-x.
- [35] G. Mensa, R. Correale, Device for controlling a gaseous flow and systems and methods employing the device (May 2017). URL https://patents.google.com/patent/US20170130870A1/en? oq=Patent+20170130870
- [36] G. Mensa, R. Correale, Portable electronic system for the analysis of time-variable gaseous flows (Jun 2017).
 URL https://patents.google.com/patent/US20170168030A1/en?
 oq=Patent+20170168030

- [37] G. Mensa, R. Correale, Portable electronic device for the analysis of a gaseous composition (Apr 2019).
 URL https://patents.google.com/patent/US10256084B2/en?oq= 10256084
- [38] A. Bagolini, R. Correale, A. Picciotto, M. Di Lorenzo, M. Scapinello, Mems membranes with nanoscale holes for analytical applications, Membranes 11 (2) (2021). doi:10.3390/membranes11020074.
- [39] A. Roth, Vacuum Technology, North Holland, 1990.
- [40] N. Gallagher, Savitzky-golay smoothing and differentiation filter (01 2020).
- [41] A. Savitzky, M. J. E. Golay, Smoothing and differentiation of data by simplified least squares procedures., Analytical Chemistry 36 (8) (1964) 1627–1639. doi:10.1021/ac60214a047.
- [42] Y. M. G. Costa, S. A. Silva, L. O. Teixeira, R. M. Pereira, D. Bertolini, A. S. Britto, L. S. Oliveira, G. D. C. Cavalcanti, COVID-19 detection on chest x-ray and ct scan: A review of the top-100 most cited papers, Sensors 22 (19) (2022). doi:10.3390/s22197303.
- [43] Z. Gao, W. Zhang, J. Li, J. Zhang, G. Huang, Principal component analysis-based feature selection for machine learning: A review, Expert Systems with Applications 140 (2020) 112920.
- [44] I. T. Jolliffe, Principal component analysis, Vol. 2, Wiley Online Library, 2002.

- [45] K. Kira, L. A. Rendell, The feature selection problem: Traditional methods and a new algorithm, in: Proceedings of the Tenth National Conference on Artificial Intelligence, AAAI'92, AAAI Press, 1992, pp. 129–134.
- [46] C. S. Greene, N. M. Penrod, J. Kiralis, J. H. Moore, Spatially uniform relieff (SURF) for computationally-efficient filtering of gene-gene interactions, BioData Min. 2 (1) (2009) 5. doi:10.1186/1756-0381-2-5.
- [47] C. S. Greene, D. S. Himmelstein, J. Kiralis, J. H. Moore, The informative extremes: Using both nearest and farthest individuals can improve relief algorithms in the domain of human genetics, Lecture Notes in Computer Science, Springer, 2010, pp. 182–193.
- [48] L. van der Maaten, G. Hinton, Visualizing data using t-SNE, Journal of Machine Learning Research 9 (86) (2008) 2579–2605.
 URL http://jmlr.org/papers/v9/vandermaaten08a.html